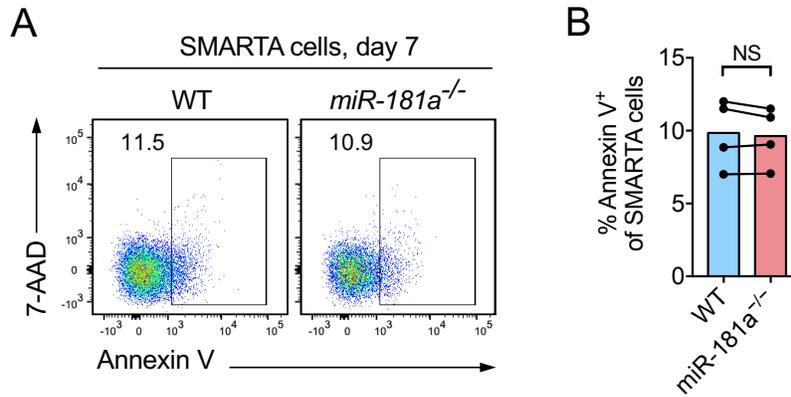


Supplemental Materials

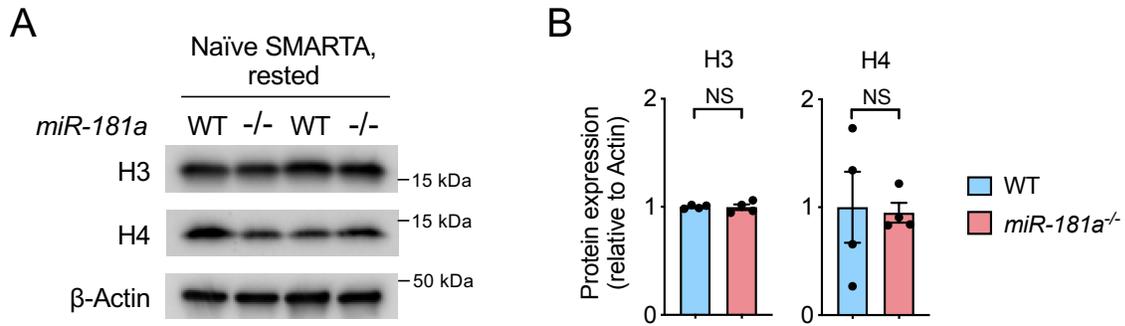
Histone deficiency and accelerated replication stress in T cell aging

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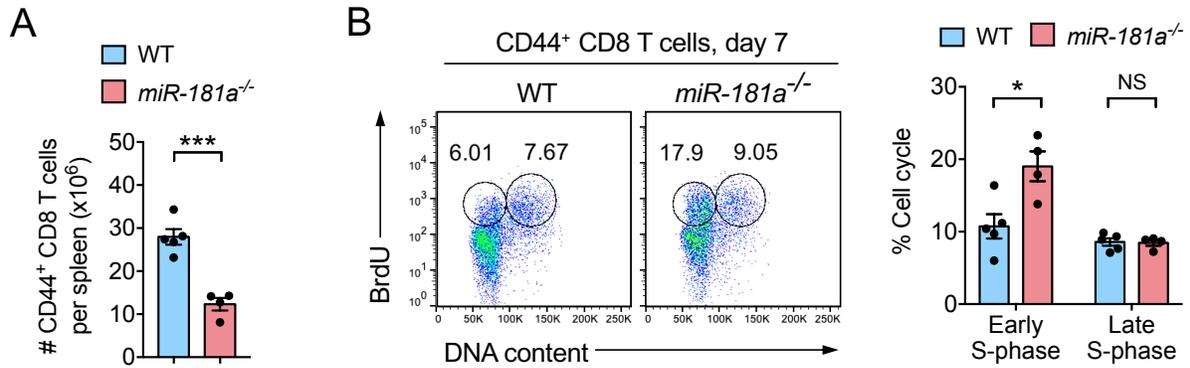
Wenqiang Cao, Lu Tian, Cornelia M. Weyand, Jörg J. Goronzy



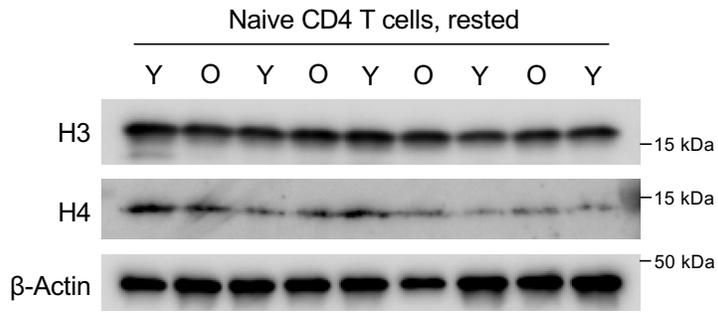
Supplemental Figure 1. Reduced expansion of miR-181a-deficient T cells is not due to increased cell death. Related to Figure 1A. Congenically marked WT and *miR-181a*^{-/-} SMARTA cells were co-transferred into B6 mice before LCMV infection. **(A)** Representative flow plots of Annexin V and 7-AAD staining of SMARTA cells on day 7 after LCMV infection. **(B)** Summary data of the percentages of Annexin V⁺ apoptotic cells (mean, two-tailed paired t test). Data are representative of two experiments with 4-5 mice per group. NS, not significant.



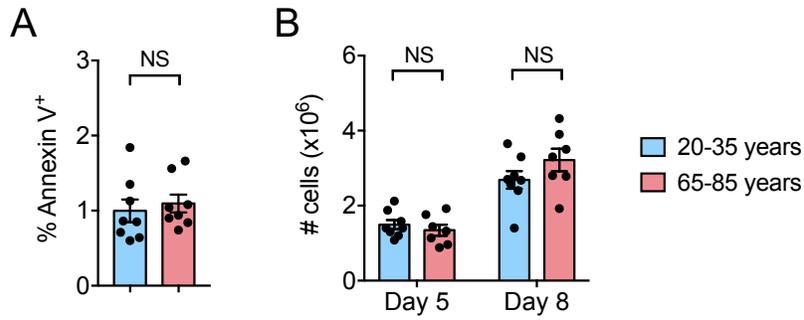
Supplemental Figure 2. Comparison of histone expression in unstimulated naïve WT and *miR-181a*-deficient T cells. Related to Figure 1D. (A) Immunoblots for histone H3 and H4 proteins from unstimulated naïve SMARTA cells. (B) Summary data of mean normalized intensities (n=4, mean \pm SEM, two-tailed unpaired t test). NS, not significant.



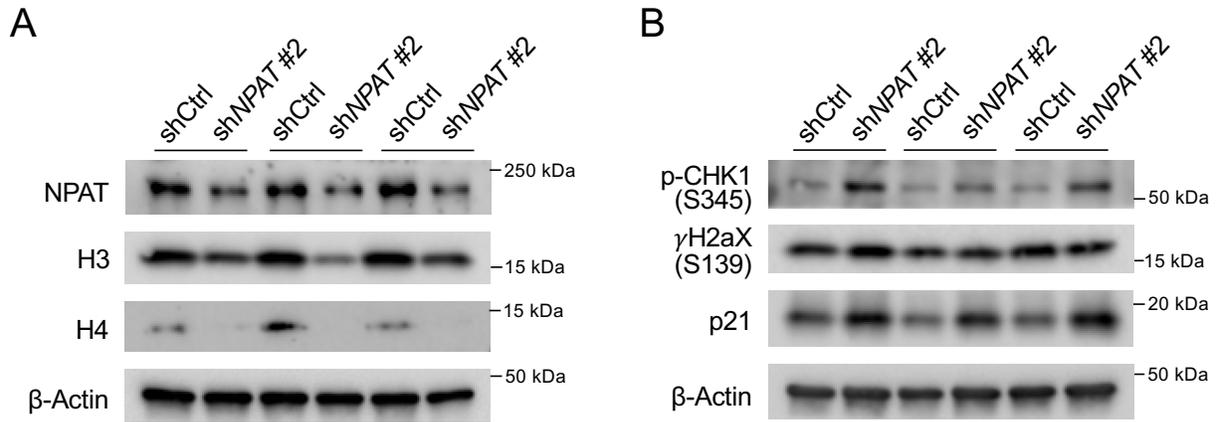
Supplemental Figure 3. Proliferative defect and accumulation of *miR-181a*^{-/-} CD8 T cells in the early S-phase of the cell cycle. Related to Figure 1, F and G. On day 7 after LCMV infection, WT and *miR-181a*^{-/-} mice were injected with BrdU for 1 hour prior to spleen harvest. **(A)** Numbers of CD44⁺ CD8 T cells (mean ± SEM, two-tailed unpaired t test). **(B)** Representative flow plots of BrdU incorporation and DNA content in CD44⁺ CD8 T cells (left) and summary of frequencies (right, mean ± SEM, two-tailed unpaired t test). Data are representative of two experiments with 3-6 mice per group. *P<0.05, ***P<0.001, NS, not significant.



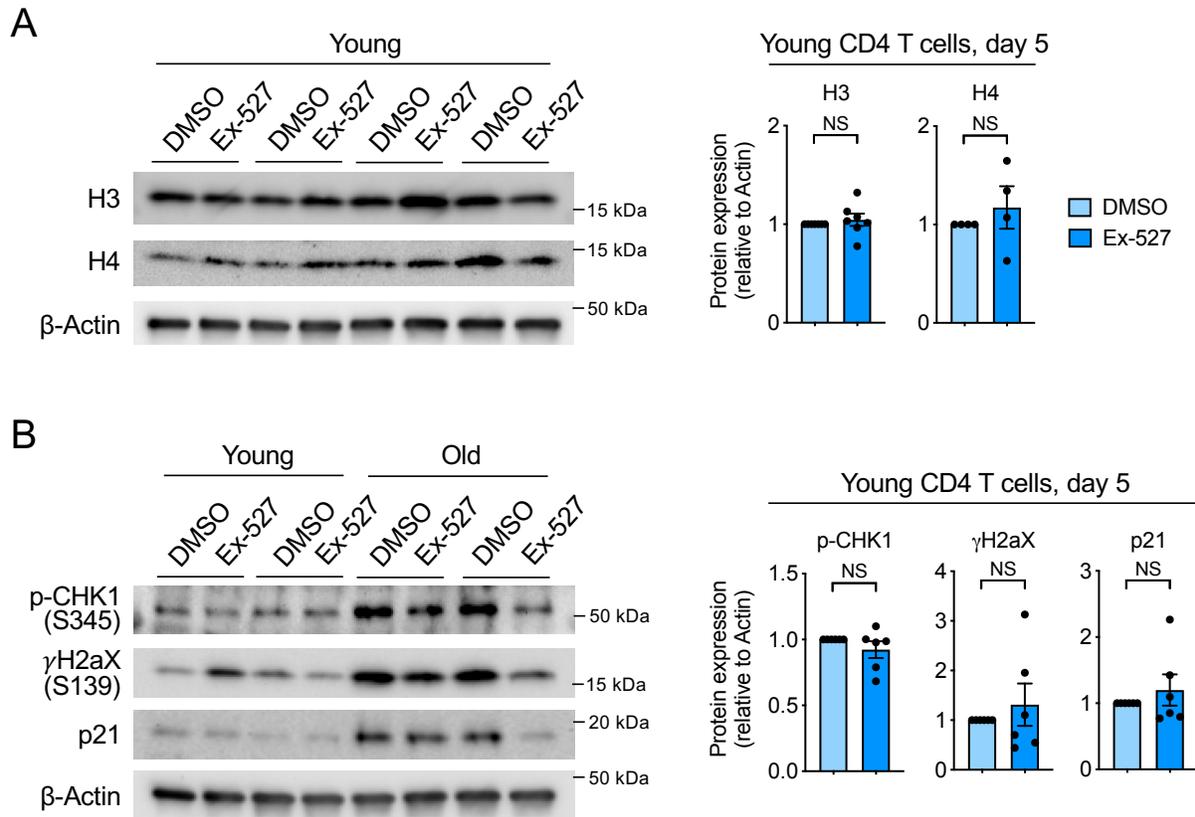
Supplemental Figure 4. Comparison of histone expression in unstimulated naïve CD4 T cells from young and old adults. Related to Figure 2C. Immunoblots for histone H3 and H4 in unstimulated naïve CD4 T cells from 5 young and 4 old individuals.



Supplemental Figure 5. Proliferative responses of naïve CD4 T cells from young and older individuals. Related to Figure 2F. Naïve CD4 T cells from healthy young and old individuals were activated with beads coated with anti-CD3/anti-CD28 antibodies. **(A)** Summary graph of frequencies of annexin V⁺ apoptotic cells on day 5 from 8 young and 8 old adults (mean ± SEM, two-tailed unpaired t test). **(B)** Summary data of the number of cells on day 5 and day 8 after activation from 8 young and 7 old individuals (mean ± SEM, two-way ANOVA followed by Tukey's post-hoc test). NS, not significant.

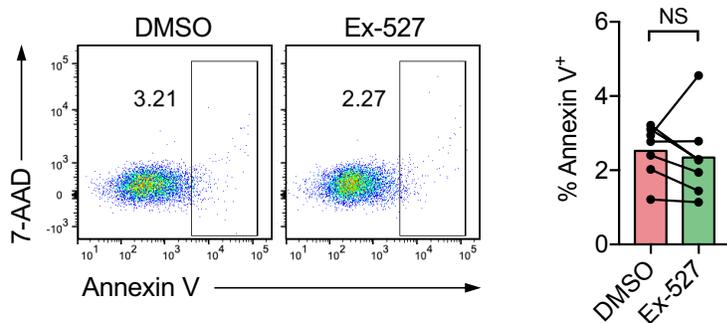


Supplemental Figure 6. Reduced histone expression promotes replication stress. Related to Figure 3. Naïve CD4 T cells from three young adults were activated with anti-CD3/anti-CD28 beads and transduced with control shRNA (shCtrl) or *NPAT* shRNA (sh*NPAT* #2) lentivirus for 6 days. Immunoblots for NPAT, histone H3 and H4 (**A**) and p-CHK1 (S345), γ H2aX (S139) and p21 (**B**) in lentivirally-transduced GFP⁺ cells.



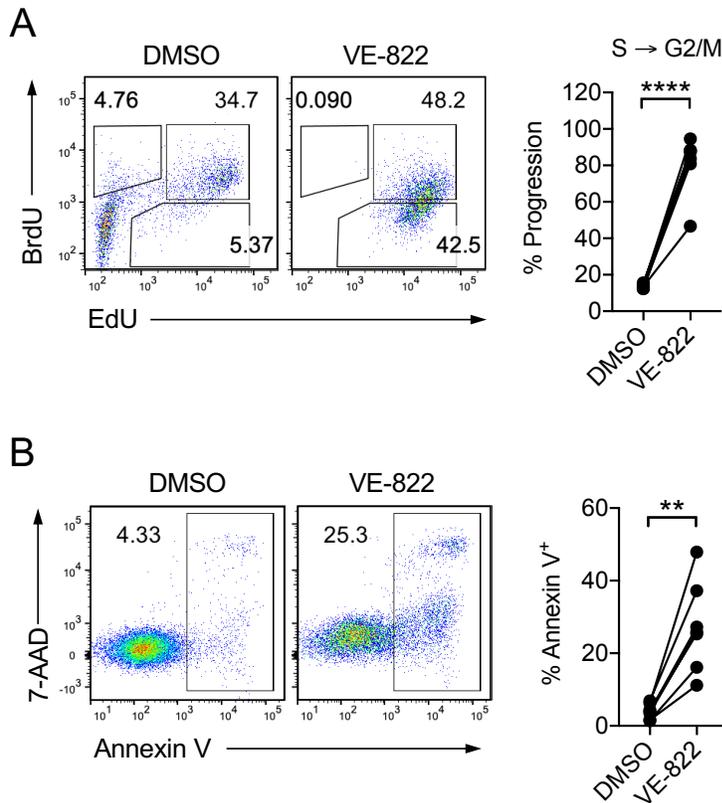
Supplemental Figure 7. Inhibition of SIRT1 activity in T cell response of young adults.

Naïve CD4 T cells from young or older individuals were activated with either DMSO or Ex-527 for 5 days. **(A)** Immunoblots of histone H3 and H4 in cycling cells (left) and summary data from 4-7 young adults (right, mean \pm SEM, two-tailed paired t test). Related to Figure 5G. **(B)** Immunoblots of indicated proteins in cycling young and old cells (left) and summary data from 6 young adults (mean \pm SEM, two-tailed paired t test). Related to Figure 6B. NS, not significant.



Supplemental Figure 8. Cell survival after inhibition of SIRT1 activity during T cell

activation. Related to Figure 6F. Naïve CD4 T cells from six old individuals were activated with anti-CD3/CD28 beads for 5 days in the presence of either DMSO or Ex-527. Representative flow plots of Annexin V and 7-AAD staining (left) and summary data of the percentages of Annexin V⁺ apoptotic cells (right, mean, two-tailed paired t test). NS, not significant.



Supplemental Figure 9. Abrogation of ATR checkpoint accelerates cell cycle progression

and increases cell death. Related to Figure 6. Naïve CD4 T cells from six old individuals were

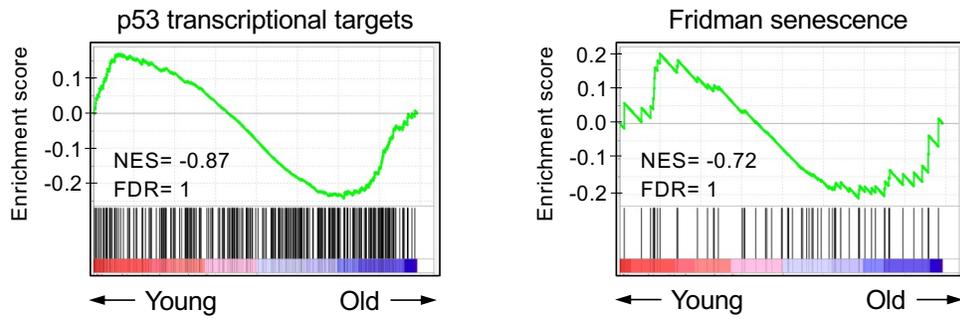
activated with anti-CD3/CD28 beads for 5 days. The ATR inhibitor VE-822 was added to the

culture on day 4. **(A)** On day 5, cells were pulsed with EdU for 2 hours, followed by BrdU for 1

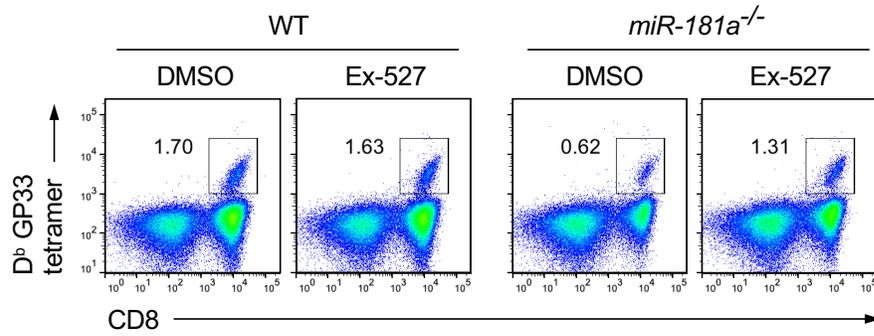
hour. Representative flow plots of BrdU and EdU incorporation (left) and summary graphs of the percentages of BrdU⁻EdU⁺ G2/M-phase cells among EdU⁺ cells (right, two-tailed paired t test).

(B) Representative flow plots of Annexin V and 7-AAD staining (left) and summary data of the

percentages of Annexin V⁺ apoptotic cells (right, two-tailed paired t test). **P<0.01, ****P<0.0001.



Supplemental Figure 10. GSEA of the transcriptome of unstimulated naïve CD4 T cells from young and old individuals. Related to Figure 7A. GSEA of p53 transcriptional target genes and genes related to senescence in the transcriptome of unstimulated young naïve CD4 T cells relative to the expression in old naïve CD4 T cells (accession number SRA: PRJNA638216).



Supplemental Figure 11. SIRT1 inhibition augments antigen-specific T cell responses in vivo. Related to Figure 8D. WT and *miR-181a*^{-/-} mice were infected with LCMV and given DMSO or Ex-527 daily starting the day after the infection. Representative flow plots of splenic D^b LCMV GP33 tetramer⁺ CD8 T cells on day 7. Summary data of number of tetramer⁺ CD8 T cells is shown in Figure 8D. Data are representative of two experiments with 5 mice per group.

Supplemental Table 1. Oligonucleotide primer sets used in this study.

Gene	Forward	Reverse
<i>HIST1H1B</i>	CTAAGGAGCGCAATGGCCTTT	CTTCGGAGTCTTCTTCACTGC
<i>HIST1H2AE</i>	CTACTCCGAACGAGTCGGG	GATGGTCACGCGACCTAGAAG
<i>HIST2H2AB</i>	CCATCTGCAACTAGCCGTGAG	CAGGCTTGTGACTCTCCGT
<i>HIST1H2BF</i>	ACCTGCTAAGTCCGCTCCT	CTACGCTTGCCTTCTTACCA
<i>HIST1H3A</i>	ACTGCTCGGAAGTCTACTGGT	GCGCTGGAAAGGTAGTTTACGA
<i>HIST1H3B</i>	ATGGCTCGTACTAAACAGACAGC	TTCCGAATCAGCAACTCGGTC
<i>HIST1H3D</i>	CCATTCCAGCGTCTAGTCCG	TCTGAAAACGCAGATCAGTCTTG
<i>HIST1H4B</i>	AAGGCGGTAAAGTTTGGGTA	GGAAATTCGCTTAACCCCACC
<i>HIST1H4E</i>	CCGTAAGGTCCTGCGAGATAA	AGTCACAGCATCACGAATCAC
<i>HIST1H4H</i>	GAGGAGCTAAGCGTCATCGC	ACACCTTCAGAACCACGAG
<i>TIPIN</i>	AGAATGGCGTGATTGACCTACC	CCAGTGCTCCATGTGTCTGATTA
<i>CDC6</i>	TGTTCTCCTCGTGAAAAGCC	GGGGAGTGTTGCATAGGTTGT
<i>CLSPN</i>	TGGAGAGTGGGGTCCATTCAT	CCGGGGTTTACGTTTGAAGAAA
<i>RFC2</i>	GTGAGCAGGCTAGAGGTCTTT	TGAGTTCCAACATGGCATCTTTG
<i>RFC4</i>	TTGGGCCTGAACTTTTCCGAT	AGCGACTTCCTGACACAGTTA
<i>RAD51</i>	CAACCCATTTACGGTTAGAGC	TTCTTTGGCGCATAGGCAACA
<i>BHLHE40</i>	GACGGGAATAAAGCGGAGC	CCGGTCACGTCTCTTTTTTCTC
<i>LIF</i>	CCAACGTGACGGACTTCCC	TACACGACTATGCGGTACAGC
<i>DDIT4</i>	TGAGGATGAACACTTGTGTGC	CCAACCTGGCTAGGCATCAGC
<i>AEN</i>	CTTCCAGGCGCTCAAGTATGT	GGGCCAGGTCCTTTAGAGAGA
<i>SDC4</i>	GGACCTCCTAGAAGGCCGATA	AGGGCCGATCATGGAGTCTT
<i>FOSL1</i>	CAGGCGGAGACTGACAACTG	TCCTTCCGGGATTTTGCAGAT
<i>CSF1</i>	TGGCGAGCAGGAGTATCAC	AGGTCTCCATCTGACTGTCAAT
<i>CSF2</i>	TCCTGAACCTGAGTAGAGACAC	TGCTGCTTGTAGTGGCTGG
<i>CCL3</i>	AGTTCTCTGCATCACTTGCTG	CGGCTTCGCTTGTTAGGAA
<i>CXCL8</i>	ACTGAGAGTGATTGAGAGTGGAC	AACCCTCTGCACCCAGTTTTTC
<i>IFIT1</i>	TTGATGACGATGAAATGCCTGA	CAGGTCACCAGACTCCTCAC
<i>IFIT2</i>	AAGCACCTCAAAGGGCAAAC	TCGGCCCATGTGATAGTAGAC
<i>MX1</i>	GTTTCCGAAGTGGACATCGCA	CTGCACAGGTTGTTCTCAGC
<i>IRF5</i>	GGGCTTCAATGGGTCAACG	GCCTTCGGTGTATTTCCCTG
<i>IRF7</i>	CCCACGCTATACCATCTACCT	GATGTCGTCATAGAGGCTGTTG
<i>RSAD2</i>	CAGCGTCAACTATCACTTCACT	AACTCTACTTTGCAGAACCTCAC
<i>ISG15</i>	CGCAGATCACCCAGAAGATCG	TTCGTGCGATTTGTCCACCA
<i>OAS3</i>	TCTGAGACTCACGTTTCCTGA	CACTGTTGAGGAGGGTAGAGTA
<i>USP18</i>	CCTGAGGCAAATCTGTCAGTC	CGAACACCTGAATCAAGGAGTTA
<i>IFI27</i>	TGCTCTCACCTCATCAGCAGT	CACAACCTCCTCAATCACAAC
<i>ACTB</i>	ATGGCCACGGCTGCTTCCAGC	CATGGTGGTGCCGCCAGACAG